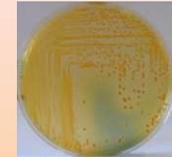


The antimicrobial effect of propolis against a range of commonly encountered human pathogens

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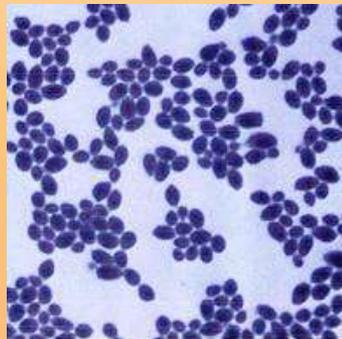
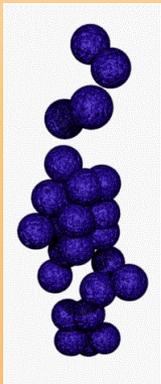
Introduction

The research carried out in this project was to assess the antimicrobial effect of propolis, against commonly isolated human pathogens using the minimum inhibitory concentrations (MIC), minimum bacterial concentration and time-kill curves.

The organisms used were control strains (NCTC) or laboratory isolates from:

- urine samples
- sputum samples
- blood cultures
- groin swabs
- high vaginal swabs

The clinical isolates used included both Gram positive and Gram negative bacteria, and yeasts.



Gram positive organisms used

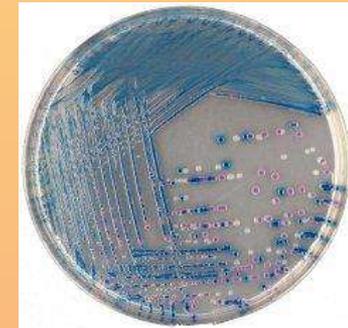
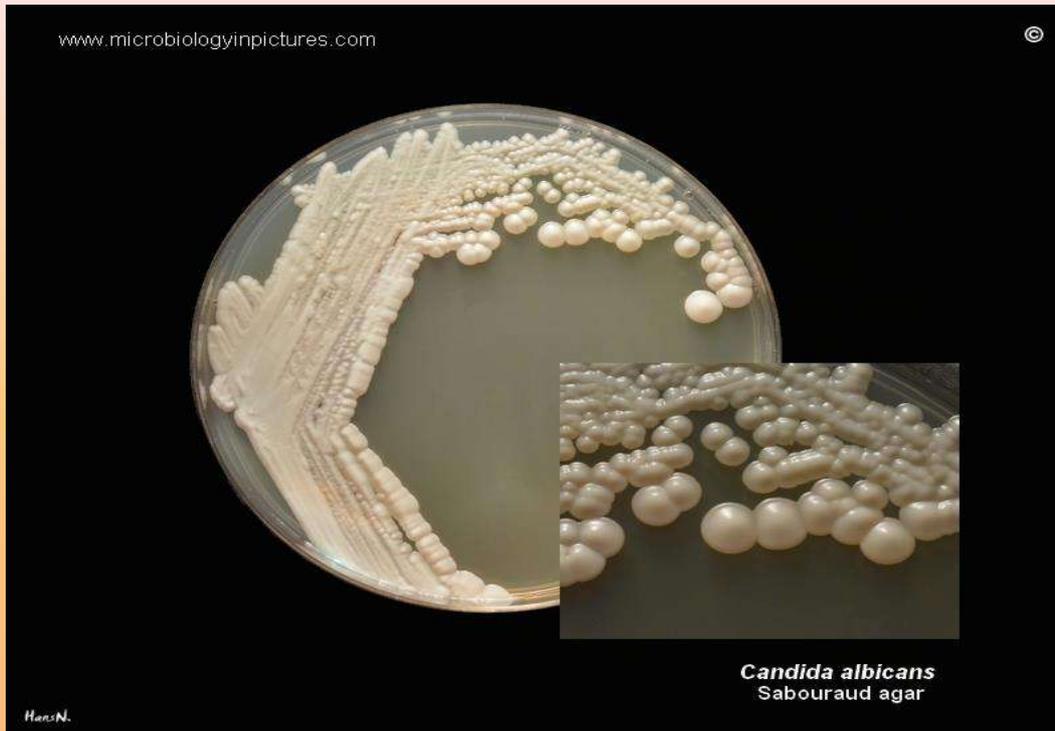
Organism	NCTC No. or isolation site
Oxford <i>Staphylococcus aureus</i>	NCTC 6571
Methicillin Resistant <i>Staphylococcus aureus</i> (MRSA)	NCTC 10443
<i>Staphylococcus auricularis</i>	Blood culture
<i>Staphylococcus lugdunensis</i>	Blood culture
Vancomycin resistant <i>Enterococcus faecium</i>	Groin swab
Methicillin Resistant <i>Staphylococcus aureus</i> (MRSA)	Blood culture
<i>Staphylococcus warneri</i>	Blood culture

Gram negative organisms used

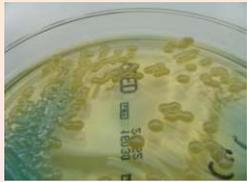
Organism	NCTC No. or isolation site
<i>Eschericia coli</i>	NCTC 10418
<i>Pseudomonas aeruginosa</i>	NCTC 10662
<i>Eschericia coli</i> (sensitive)	Urine
<i>Eschericia coli</i> (resistant)	Urine
<i>Pseudomonas aeruginosa</i>	Urine
<i>Proteus mirabilis</i> (sensitive)	Urine
<i>Proteus mirabilis</i> (resistant)	Urine
<i>Klebsiella oxytoca</i> (Ampicillin sensitive)	Urine
<i>Klebsiella oxytoca</i> (Ampicillin resistant)	Urine
<i>Enterobacter cloacae</i>	Sputum

Yeasts used

Organism	Isolation site
<i>Candida albicans</i>	High vaginal swab
<i>Candida albicans</i>	High vaginal swab
<i>Candida parapsilosis</i>	High vaginal swab
<i>Cryptococcus humicolus</i>	High vaginal swab

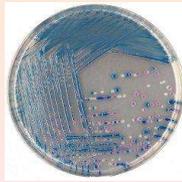


MIC determination – agar dilution method

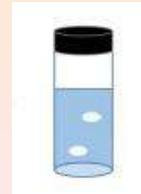


Bacterial colonies

or

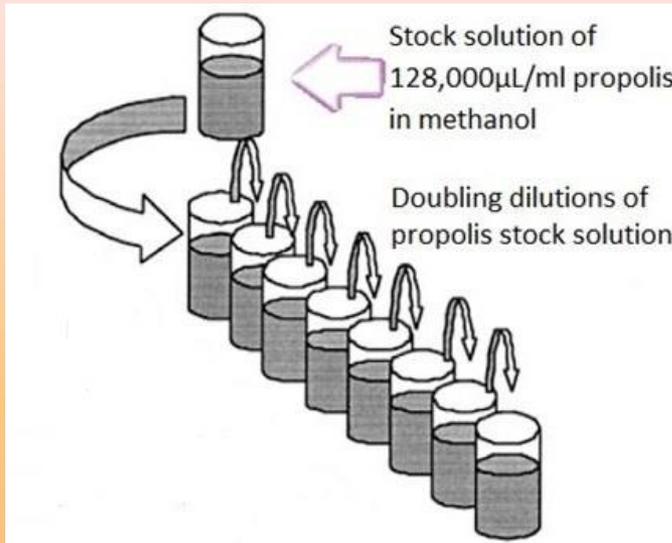


Yeast colonies



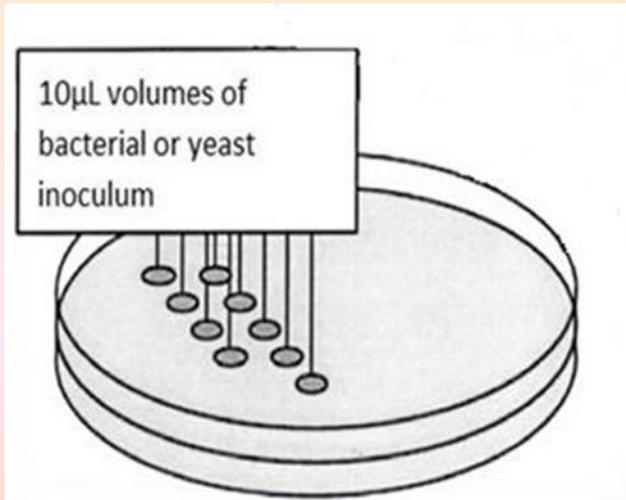
Bacteria or yeast inoculum equivalent to 0.5 MacFarlane (1.5×10^8 /ml)

1ml defibrinated horse blood and 1 ml propolis solution added to 19ml isosensitest agar at 65°C

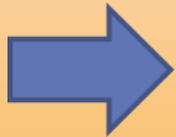


Final range of concentrations of propolis in agar: 64 mg/ml – 0.125mg/ml

MIC determination – agar dilution method



37°C + 5% CO₂
for 18 hours



MBC determination

At any given concentration of propolis, any areas showing no growth of bacteria were sub cultured onto Columbia blood agar & incubated for 18 hours to determine the minimum bactericidal concentration (MBC) of propolis for that organism.

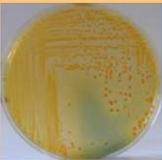
Time kill curves method

Stock solutions of propolis were made up in concentrations of:

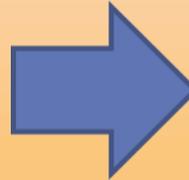


- 128mg/L
- 64mg/L
- 32mg/L
- 16mg/L
- 8mg/L
- 4mg/L
- 2mg/L
- 1mg/L
- 0.5mg/L

in sterile nutrient broth containing dimethylsulphoxide (5% v:v) and Tween 80 (20% v:v). The sterile nutrient broth was also used as a negative control.

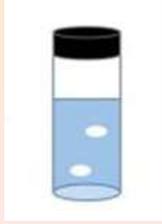


Sterile peptone water
containing the bacteria
or yeast inoculum
equivalent to 0.5
MacFarlane
(1.5×10^8 /ml)



37°C + 5% CO₂
for 18 hours

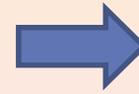
Time kill curves method



1ml



1ml



3000 rpm
10 minutes

Sterile peptone water containing the bacteria or yeast inoculum equivalent to 0.5 MacFarlane ($1.5 \times 10^8/\text{ml}$)

Propolis solution in nutrient broth with DMS & Tween 80

37°C + 5% CO₂ for 3, 6, 9, or 12 hours

Supernatant removed



10μL



10μL into sterile distilled water, two 1 in 100 dilutions with sterile distilled water.



Bacterial pellet re-suspended in peptone water for 20 minutes



37°C + 5% CO₂ for 18 hours



Viable count performed

Minimum inhibitory concentration (MIC) and minimum bactericidal concentration results (MBC) for the Gram positive organisms

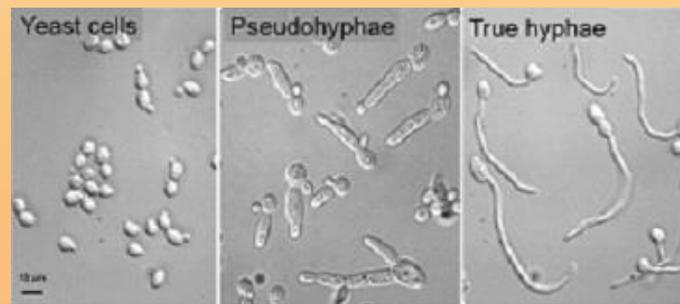
Organism	NCTC No. (if applicable)	MIC	MBC
<i>Oxford Staphylococcus aureus</i>	NCTC 6571	0.5mg/L	4mg/L
Methicillin resistant <i>Staphylococcus aureus</i> (MRSA)	NCTC 10443	0.5mg/L	4mg/L
<i>Staphylococcus auricularis</i>	n/a	0.25mg/L	2mg/L
<i>Staphylococcus lugdunensis</i>	n/a	8mg/L	64mg/L
<i>Staphylococcus warneri</i>	n/a	1mg/L	8mg/L
Methicillin resistant <i>Staphylococcus aureus</i> (MRSA)	n/a	0.5mg/L	4mg/L
Vancomycin resistant <i>Enterococcus faecium</i> (VRE)	n/a	2mg/L	8mg/L
<i>Enterococcus faecalis</i>	n/a	2mg/L	8mg/L

Minimum inhibitory concentration (MIC) and minimum bactericidal concentration results (MBC) for the Gram negative organisms

Organism	NCTC No. (if applicable)	MIC	MBC
<i>Eschericica coli</i>	NCTC 10418	64mg/L	128mg/L
<i>Eschericica coli</i> (sensitive)	n/a	64mg/L	128mg/L
<i>Eschericica coli</i> (resistant)	n/a	64mg/L	128mg/L
<i>Pseudomonas aeruginosa</i>	NCTC 10662	64mg/L	128mg/L
<i>Pseudomonas aeruginosa</i>	n/a	64mg/L	128mg/L
<i>Klebsiella oxytoca</i> (Ampicillin sensitive)	n/a	>64mg/L	>64mg/L
<i>Klebsiella oxytoca</i> (Ampicillin resistant)	n/a	>64mg/L	>64mg/L
<i>Proteus mirabilis</i> (sensitive)	n/a	>64mg/L	>64mg/L
<i>Proteus mirabilis</i> (resistant)	n/a	>64mg/L	>64mg/L
<i>Enterobacter cloacae</i>	n/a	>64mg/L	>64mg/L

Minimum inhibitory concentration (MIC) and minimum bactericidal concentration results (MBC) for the yeasts

Organism	MIC	MBC
<i>Candida albicans</i>	8mg/L	16mg/L
<i>Candida albicans</i>	8mg/L	64mg/L
<i>Candida parapsilosis</i>	16mg/L	32mg/L
<i>Cryptococcus humicolus</i>	2mg/L	8mg/L



Results – Control Organism

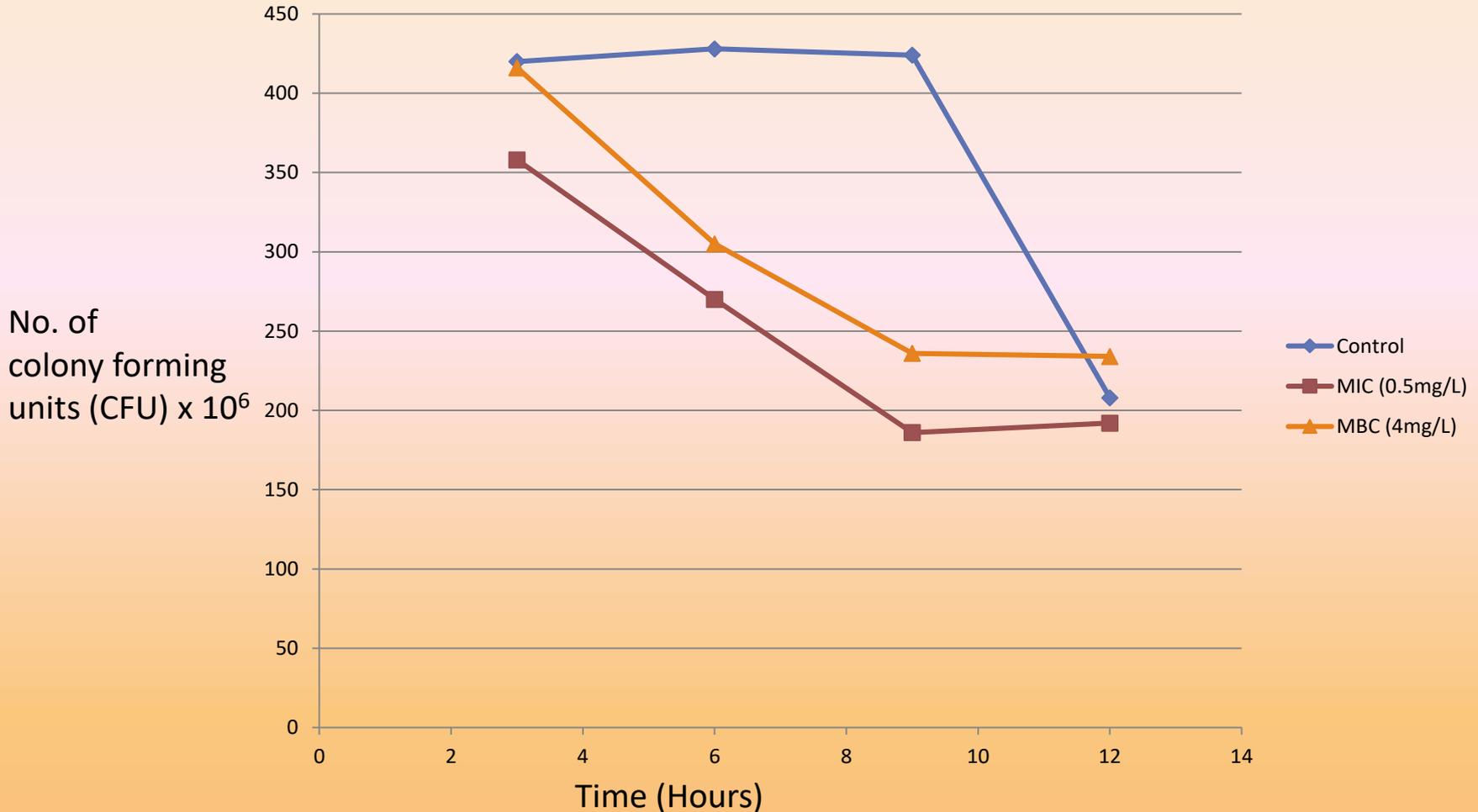


Figure 1: Time kill curves for Oxford *Staphylococcus aureus* (NCTC 6571) at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Control organism

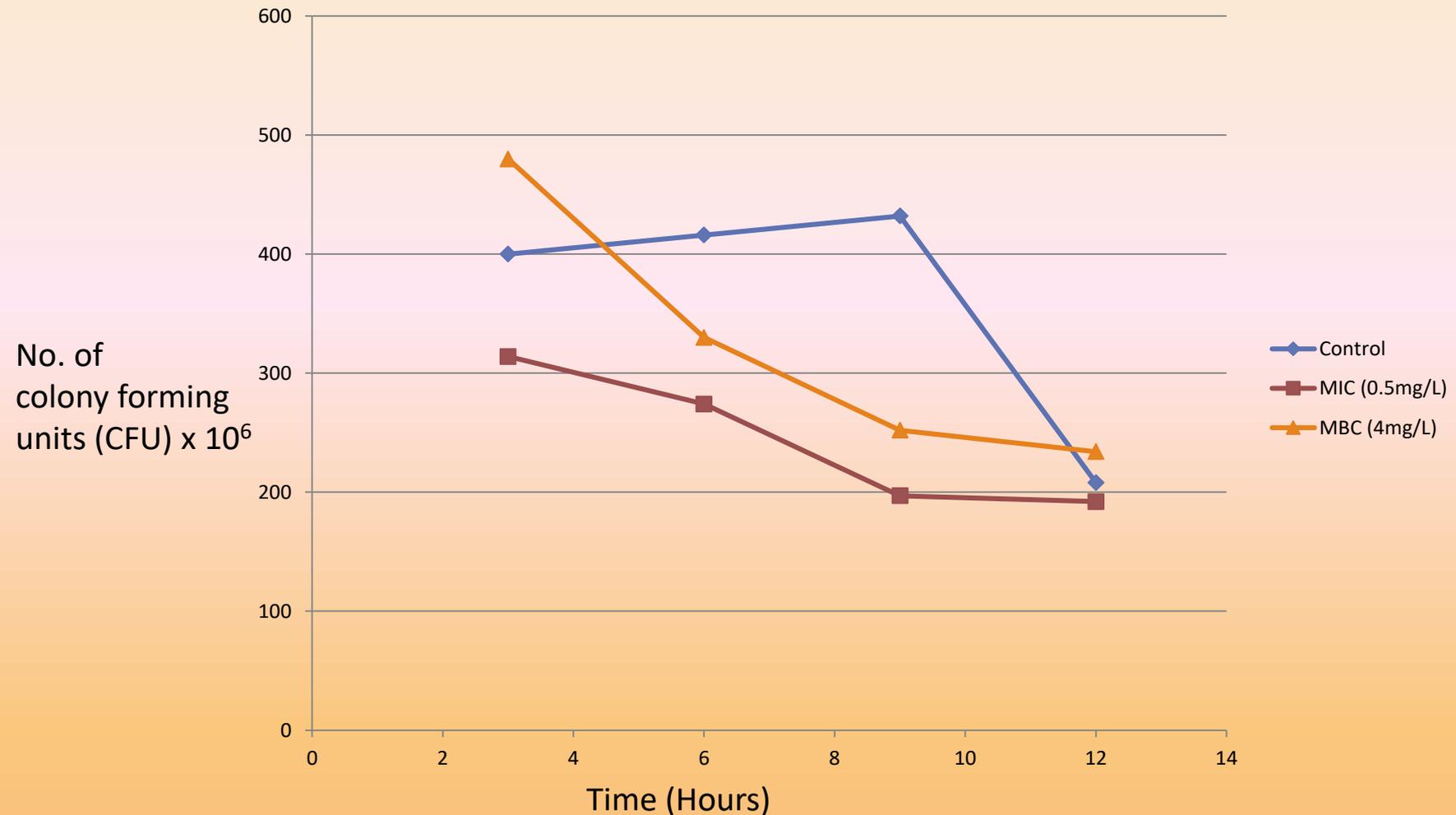


Figure 2: Time kill curves for Methicillin Resistant *Staphylococcus aureus* (NCTC 10443) at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Clinical isolate

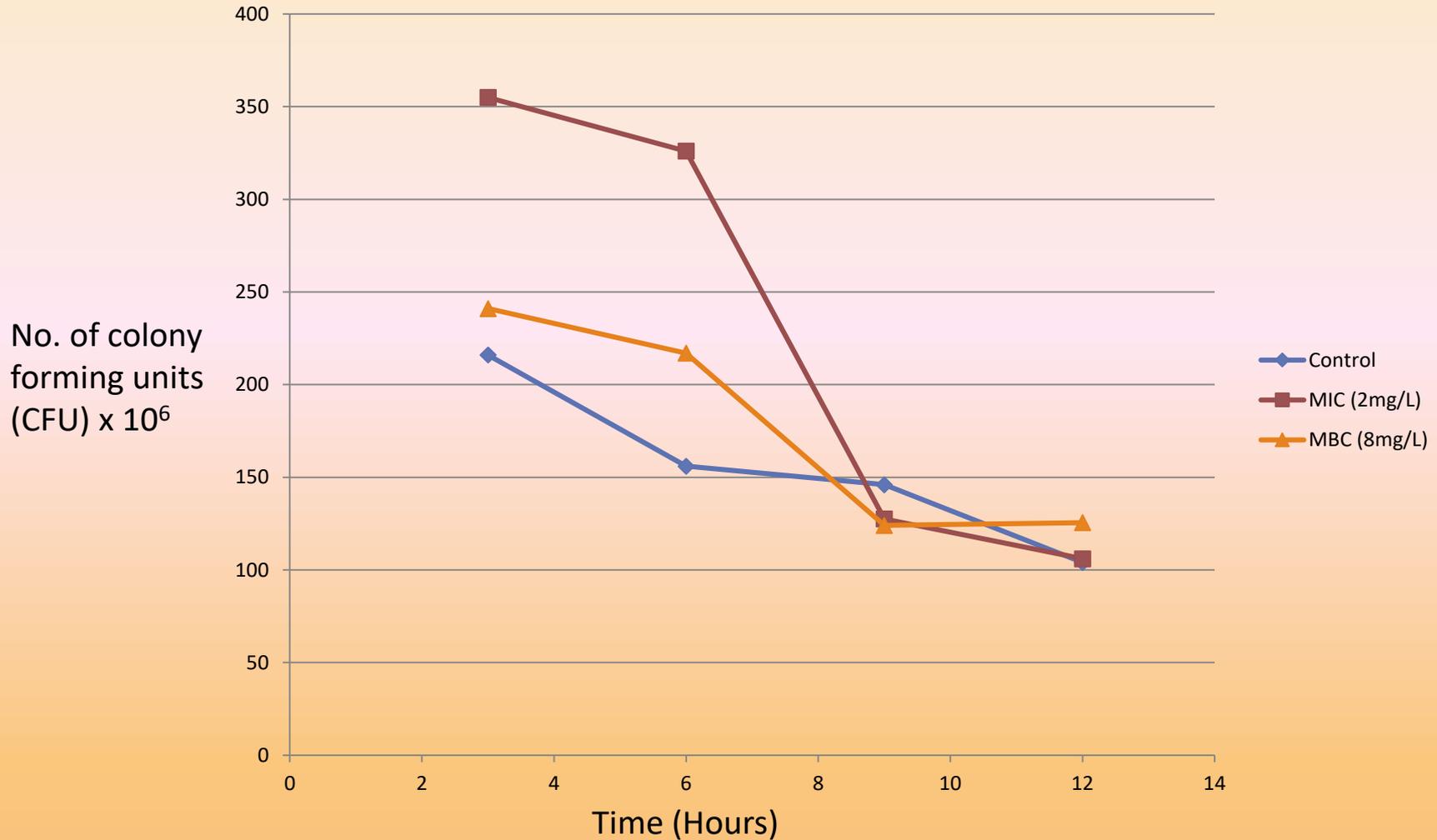


Figure 3: Time kill curves for Vancomycin resistant *Enterococcus faecium* (VRE) at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Clinical isolate

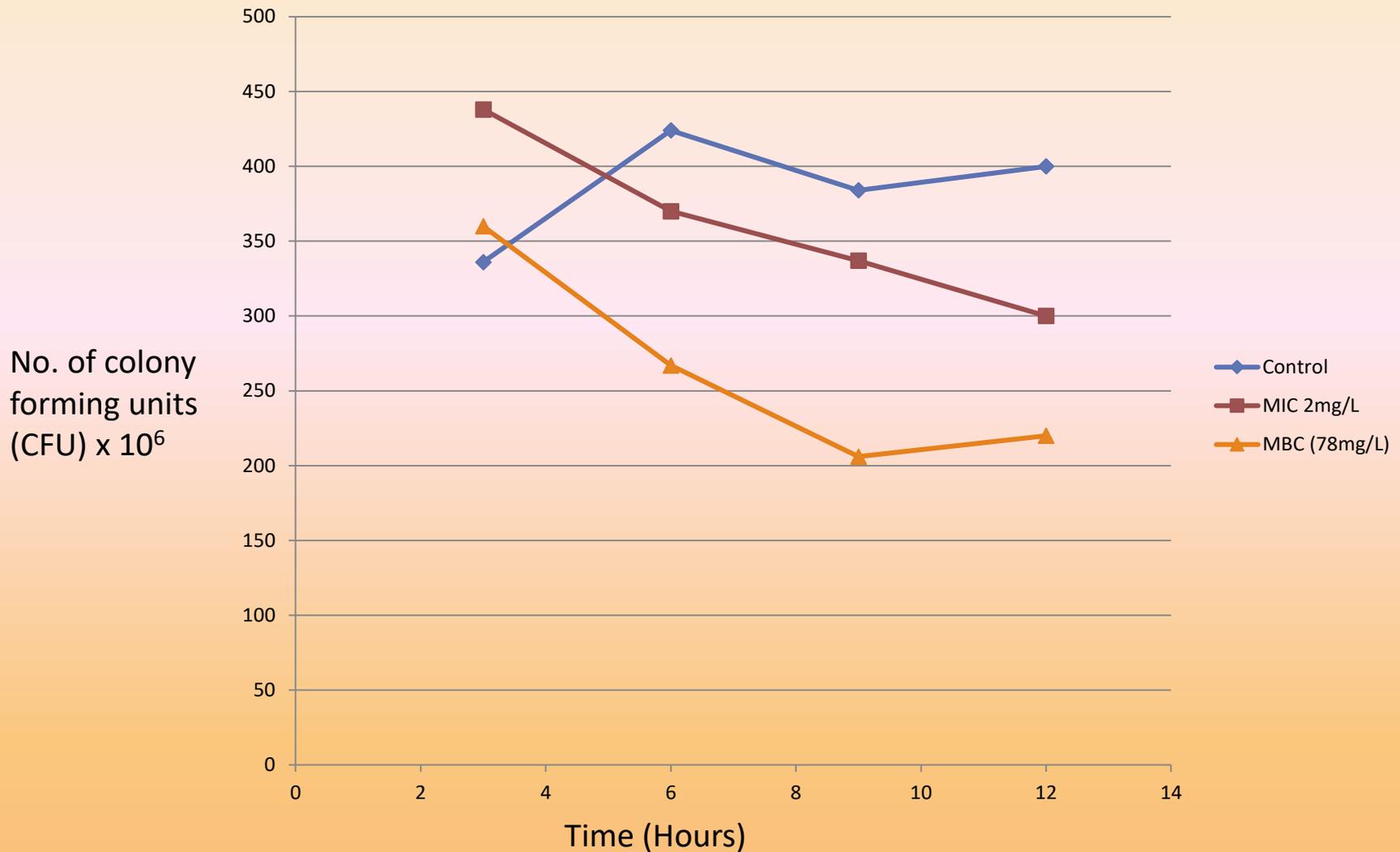


Figure 4: Time kill curves for *Enterococcus faecalis* at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Clinical isolate

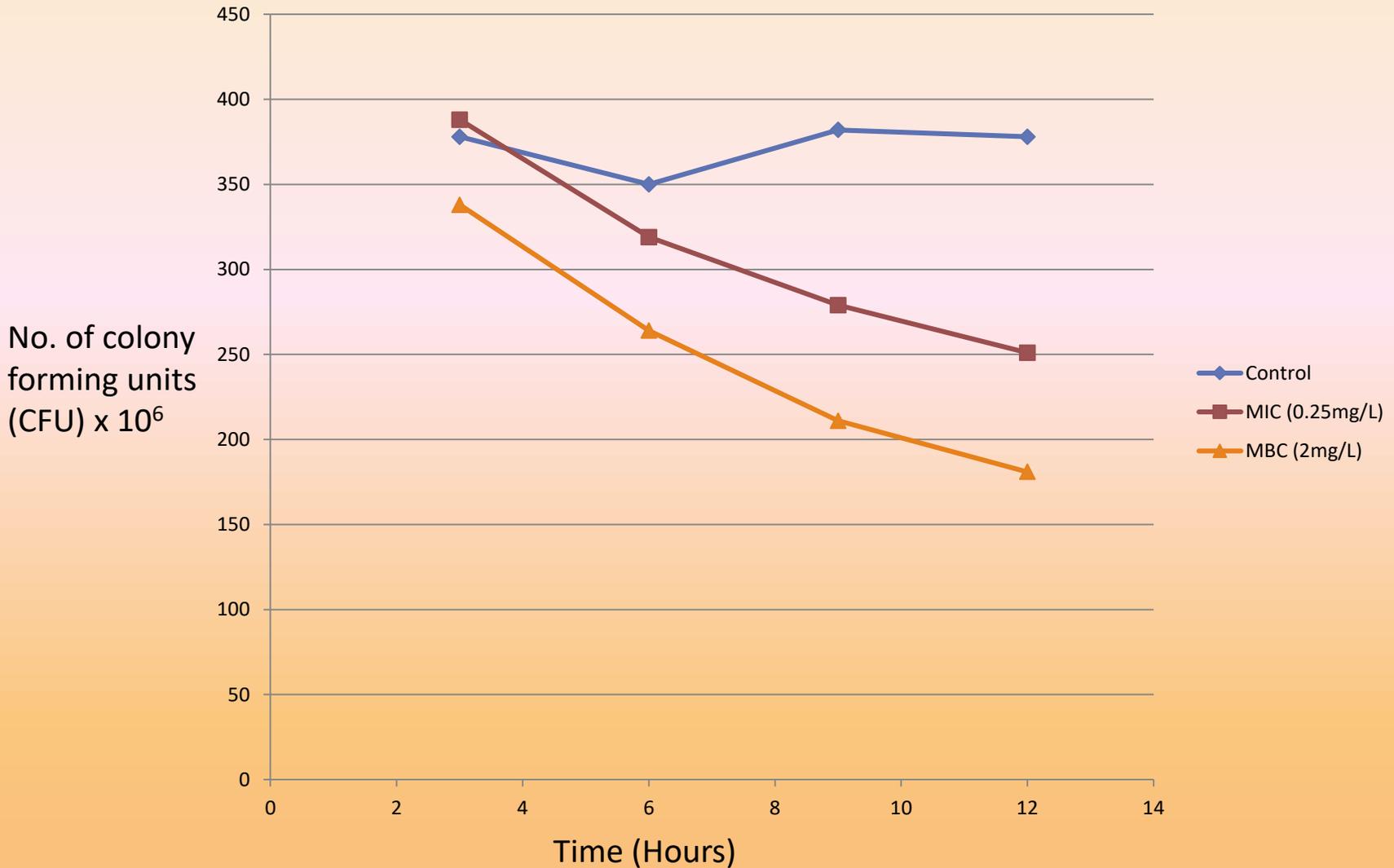


Figure 6: Time kill curves for *Staphylococcus auricularis* at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Control organism

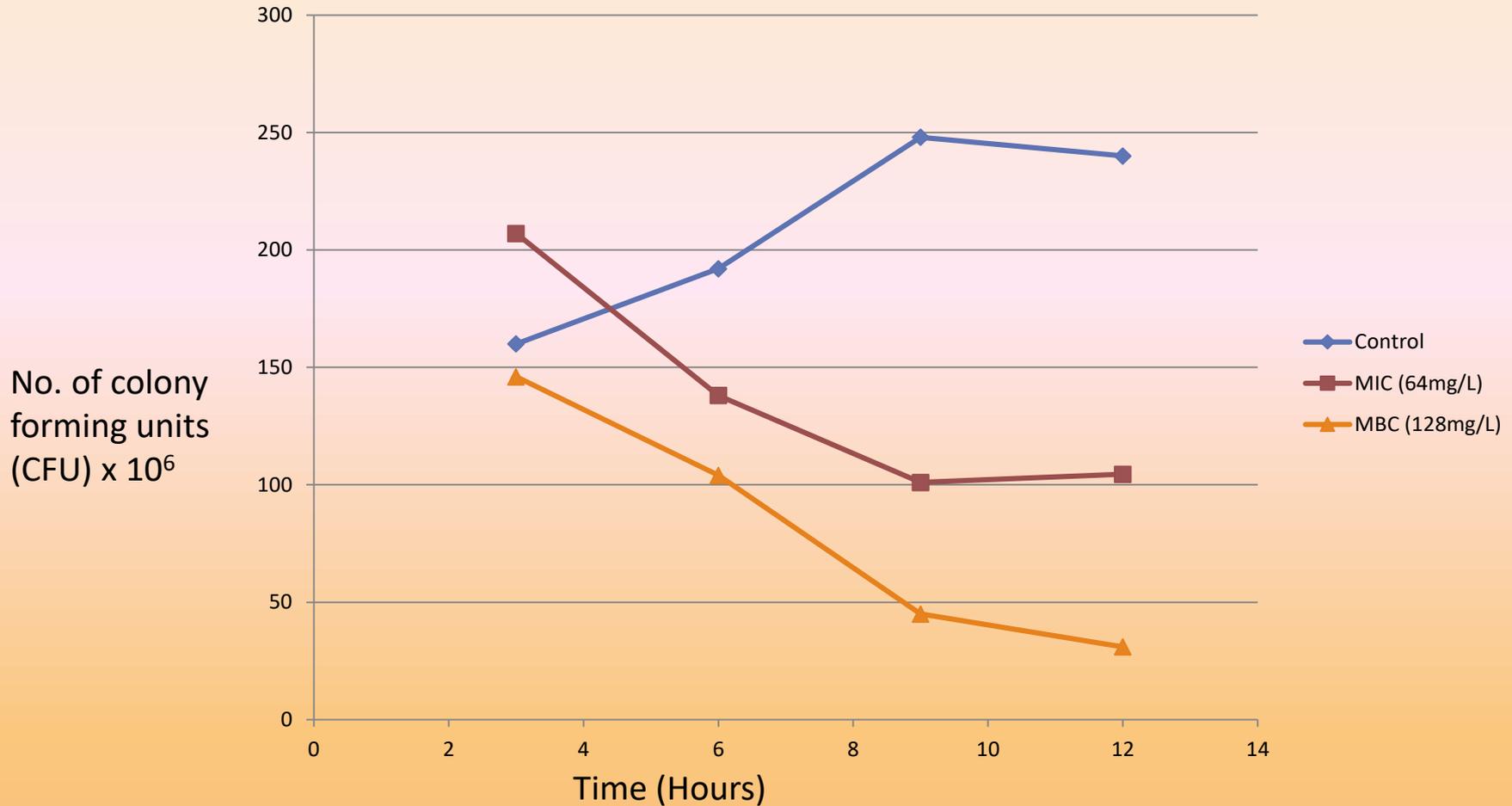


Figure 5: Time kill curves for *Escherichia coli* (NCTC 10418) at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results - Control organism

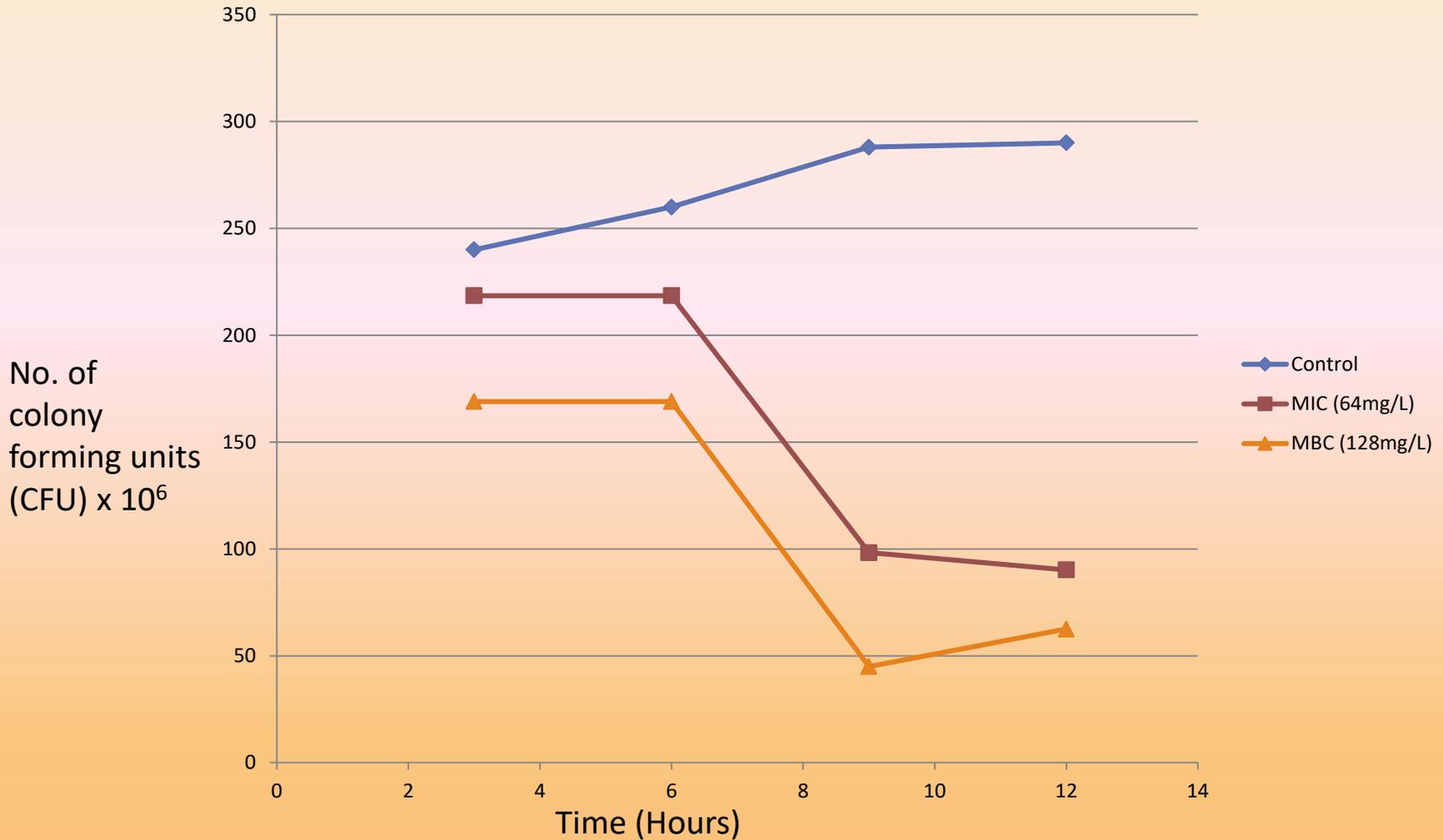


Figure 6: Time kill curves for *Pseudomonas aeruginosa* (NCTC 10662) at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Clinical isolate

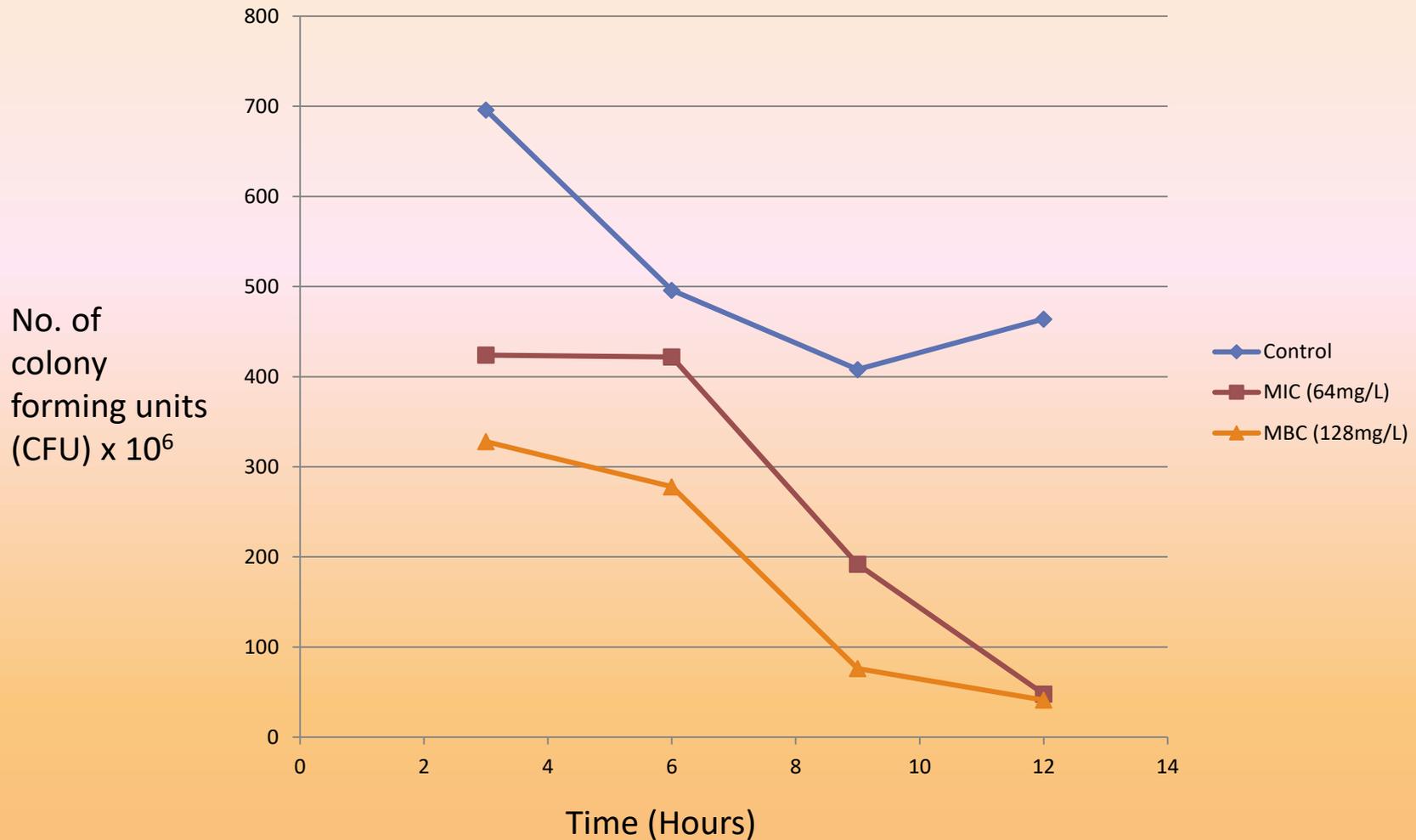


Figure 7: Time kill curves for *Escherichia coli* at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Clinical isolate

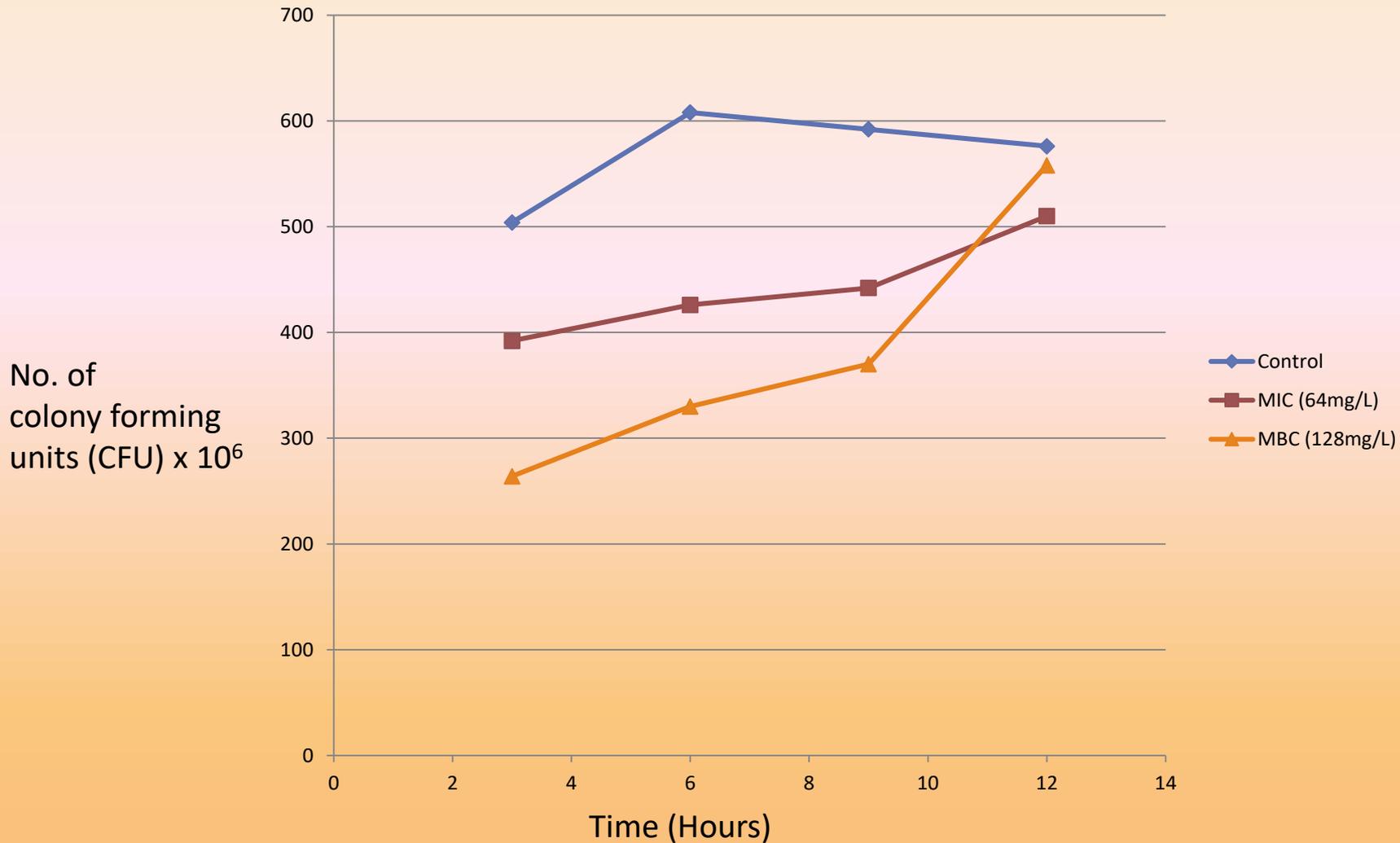


Figure 8: Time kill curves for *Klebsiella oxytoca* at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

Results – Clinical isolate

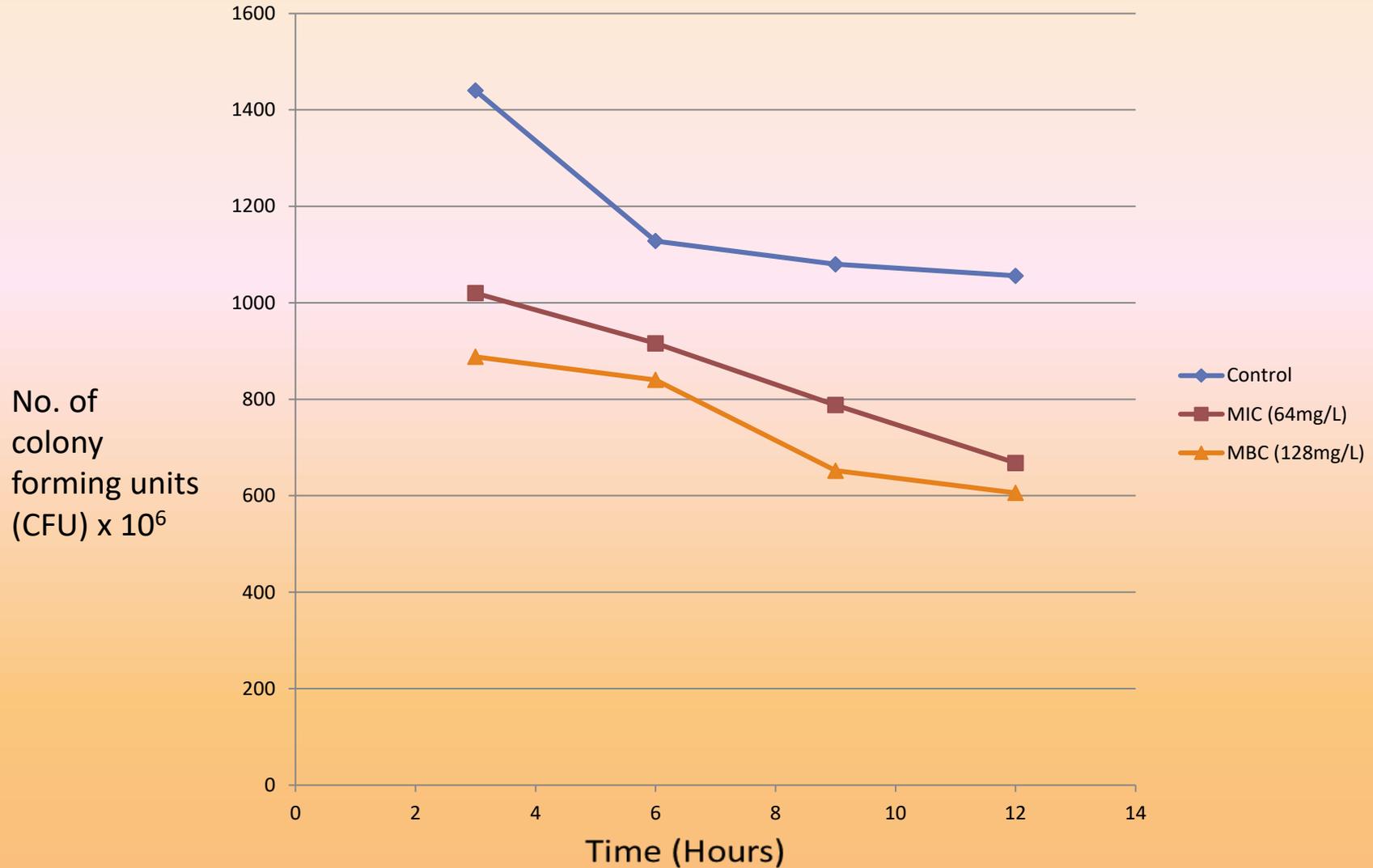


Figure 9: Time kill curves for *Enterobacter cloacae* at concentrations of propolis equivalent to the minimum inhibitory concentration (MIC) & minimum bactericidal concentration (MBC)

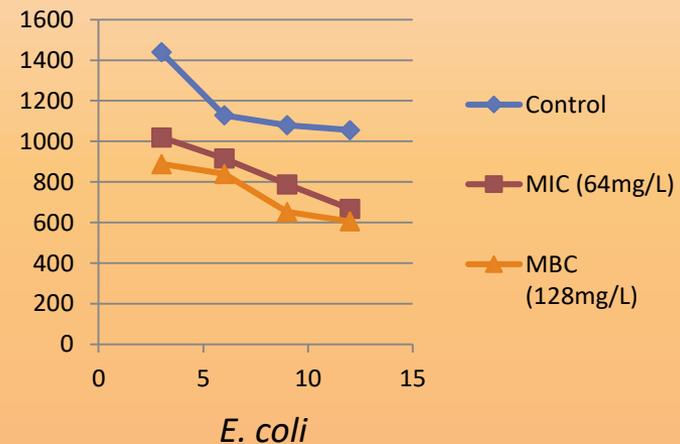
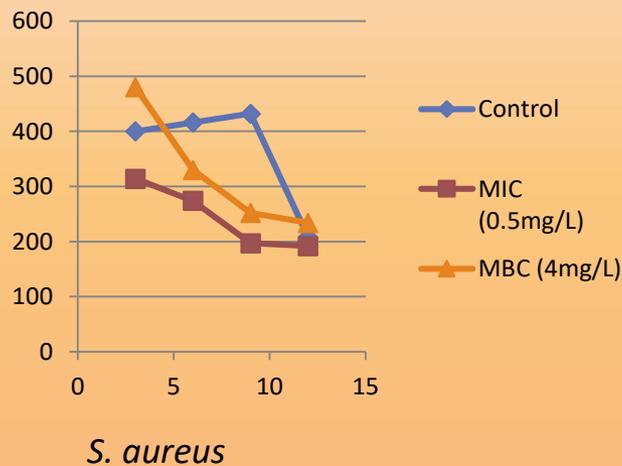
Summary of results

- The MIC and MBC investigations show that propolis has both antimicrobial and anti-fungal activity.
- Particularly effective against Gram positive organisms, such as staphylococci and streptococci, and yeasts.
- Less effective against Gram negative organisms.



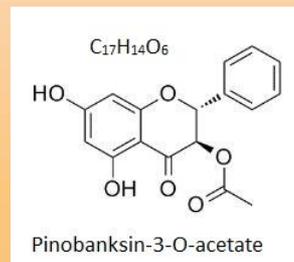
Time-kill curves:

- At lower concentrations propolis is bacteriostatic against Gram positive organisms.
- At higher concentrations, propolis is bactericidal against both Gram negative and Gram positive organisms.



Flavonoids in propolis and their activity

Organisms	Active flavinoid	Reference
<i>S. aureus</i>	Phenolic compounds (i.e. flavinoids)	Ristivojević <i>et al</i> (2016) PLoS ONE 11 (16): e0157097 Doi: 10.1371/journal.pone.0157097
<i>S.aureus, E. coli & C. albicans</i>	Pinocembrin	Rasul <i>et al</i> (2013) BioMed Research International, Article ID 379850
MRSA & <i>S. aureus</i>	Pinocembrin, Pinobankin-3-O-acetate & Chrysin	Darwish <i>et al</i> (2010) African Journal of Biotechnology Vol. 9 (36): 5966-5974
<i>E. faecalis, S. aureus & E. coli</i>	Pinobankin	Uzel <i>et al</i> (2005) Microbiological Research 189-195
MRSA, <i>Enterococcus spp., P. aeruginosa</i>	Galangin	S. Pepeljnjak & I. Kosalec (2004) FEMS Microbiology letters 240: 111-116

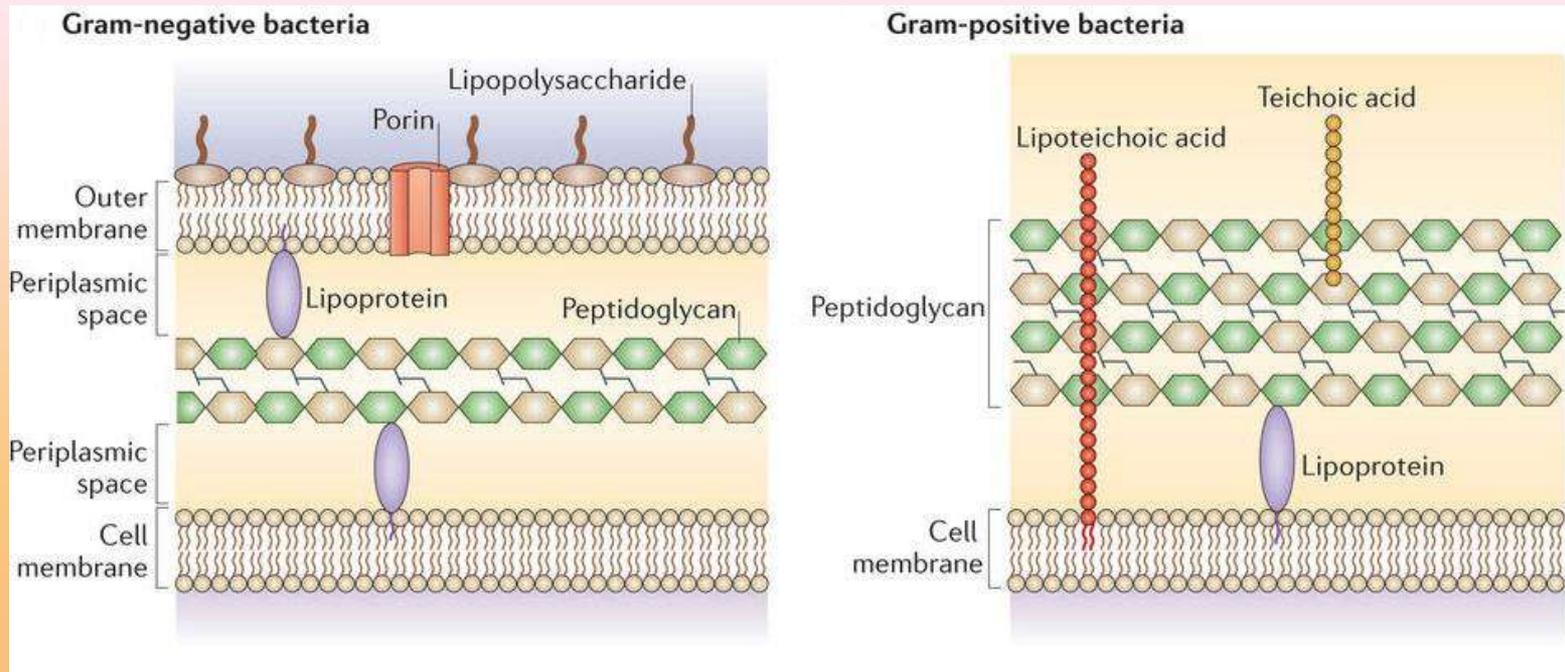


Propolis - batch analysis details:

- Chrysin (5,7-dihydroxyflavone) 3.16% w/w
- Galangin (3,5,7-trihydroxyflavone) 2.39% w/w
- Pinocembrin (5,7-dihydroxyflavone) 2.44% w/w
- Lead 0.5 mg/kg

Conclusion

- The double membrane layer of Gram negative organisms, reduces the ability of propolis to access it's site of action – the cytoplasm.
- Gram positive organisms only have a single membrane, whereas Gram negative organisms have a double membrane.
- A mucoid layer around the bacterial cells in some Gram negative organisms, such as *E. cloacae* & *K. oxytoca*, also hinders the action of propolis.



Overall, both the antimicrobial activity of propolis, and it's possible clinical applications merits further investigation.

Any questions?



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